Table 1 Inspection and Treatment of SPF Cattle Herds

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Aino virus | JaNAr28 | Every three months | Either five animals per group or 10% of the animals, whichever is greater | SN | All the antibody-positive group and cohabitant group3) culling |
| Kasbah virus | K-47 | 〃 | 〃 | SN | 〃 |
| Bovine adenovirus | Fukuroi | 〃 | 〃 | HI | 〃 |
| Japanese encephalitis virus | Nakayama | 〃 | 〃 | HI | 〃 |
| Bovine enterovirus |  | 〃 | 〃 | SN | 〃 |
| Bovine rhinovirus |  | 〃 | 〃 | SN | 〃 |
| Bovine reovirus |  | 〃 | 〃 | HI | 〃 |
| Ibaraki virus | No.2 | 〃 | 〃 | HI | 〃 |
| Akabane virus | JaGAr39 OBE-1 | 〃 | 〃 | HI ELISA | 〃 |
| Aujeszky's disease virus |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| Bluetongue virus |  | 〃 | 〃 | AGP | All the antibody-positive group and cohabitant  groupculling |
| Bovine coronavirus | Kakegawa | 〃 | 〃 | HI | 〃 |
| Bovine ephemeral fever virus | YHL | 〃 | 〃 | SN | 〃 |
| Infectious bovine rhinotracheitis virus | No.758 | 〃 | 〃 | SN | 〃 |
| Bovine leukemia virus | FLK-HK-11 FLK-SP | 〃 | 〃 | AGP, passive HI | 〃 |
| Bovine papillomavirus |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| Bovine parvovirus | BF-15 | 〃 | 〃 | HI | All the antibody-positive group and cohabitant  groupculling |
| Bovine papular stomatitis virus - pseudocowpox virus |  | 〃 | 〃 | AGP | 〃 |
| Bovine respiratory syncytial virus | NMK7 | 〃 | 〃 | SN | 〃 |
| Rotavirus | Lincoln | 〃 | 〃 | SN | 〃 |
| Bovine viral diarrhea virus | Nose | 〃 | 〃 | SN | 〃 |
| Malignant catarrhal fever virus | Cattle/elk type | 〃 | 〃 | IFA | 〃 |
| Parainfluenza virus type 3 | BN-1 | 〃 | 〃 | HI | 〃 |
| *Coxiella burneti*i |  | 〃 | 〃 | IFA | 〃 |
| *Mycoplasma* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Actinomyces bovis* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Mannheimia haemolytica* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Pasteurella multocida* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Histophilus somni* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Bacillus anthracis* |  | 〃 | 〃 | Clinical symptoms | 〃 |
| *Brucella* | *Melitensis* | 〃 | 〃 | AGG CF | All the antibody-positive group and cohabitant groupculling |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Mycobacterium bovis, Mycobacterium tuberculosis* | Aoyama B and 10 | 〃 | 〃 | Tuberculin reaction | All the antibody-positive group and cohabitant group culling |
| *Mycobacterium avium subsp. paratuberculosis* | *Mycobacterium avium* | 〃 | 〃 | Johnin reaction, ELISA | 〃 |
| *Leptospira interrogans* | *Hebdomadis* | 〃 | 〃 | AGG | 〃 |
| Piroplasma |  | 〃 | 〃 | Blood test | All the positive group and cohabitant group culling |
| Anaplasma | Marginale | 〃 | 〃 | CF | All the antibody-positive group and cohabitant groupculling |
| Epidemic hemorrhagic disease virus4) |  |  |  |  |  |
| Cowpox virus, Vaccinia virus4) |  |  |  |  |  |
| Foot and mouth disease virus4) |  |  |  |  |  |
| Lumpy skin disease virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |
| Rift Valley fever virus4) |  |  |  |  |  |
| Rinderpest virus4) |  |  |  |  |  |
| Vesicular stomatitis virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of cattle shall be completely recorded. For dead cattle, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, SN: Serum neutralization test, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, AGP: Agar gel precipitation test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 2 Inspection and Treatment of SPF Sheep Flocks

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Rotavirus | Tochigi | Every three months | Either five animals per group or 10% of the animals, whichever is greater | SN | All the antibody-positive group and cohabitant  group 3)culling |
| Bovine adenovirus | Fukuroi | 〃 | 〃 | SN | 〃 |
| Akabane virus | JaGAr39 | 〃 | 〃 | SN | 〃 |
| Aujeszky's disease virus |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| Bluetongue virus |  | 〃 | 〃 | AGP | All the antibody-positive group and cohabitant group culling |
| Infectious bovine rhinotracheitis virus | No.758 | 〃 | 〃 | SN | 〃 |
| Bovine leukemia virus | FLK | 〃 | 〃 | AGP | 〃 |
| Bovine papillomavirus |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| Bovine viral diarrhea virus | Nose | 〃 | 〃 | SN | All the antibody-positive group and cohabitant group culling |
| Parainfluenza virus type 3 | BN-1 | 〃 | 〃 | HI | 〃 |
| Scrapie |  | 〃 | 〃 | Clinical symptoms5) | All the positive group and cohabitant group culling |
| Contagious pustular dermatitis virus |  | 〃 | 〃 | SN | All the antibody-positive group and cohabitant group culling |
| Visna-maedi virus |  | 〃 | 〃 | SN | 〃 |
| Japanese encephalitis virus | Nakayama | 〃 | 〃 | HI | 〃 |
| Malignant catarrhal fever | Cattle/elk type | 〃 | 〃 | IFA | 〃 |
| *Mycoplasma* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Mycobacterium bovis, Mycobacterium tuberculosis* |  | 〃 | 〃 | Tuberculin reaction | All the antibody-positive group and cohabitant group culling |
| *Mycobacterium avium subsp. paratuberculosis* |  | 〃 | 〃 | Johnin reaction | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Brucella* |  | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Pasteurella multocida* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| Epidemic hemorrhagic disease virus4) |  |  |  |  |  |
| Cowpox virus, Vaccinia virus4) |  |  |  |  |  |
| Foot and mouth disease virus4) |  |  |  |  |  |
| Rift valley fever virus4) |  |  |  |  |  |
| Border disease virus4) |  |  |  |  |  |
| Borna disease virus4) |  |  |  |  |  |
| Ovine pulmonary adenomatosis virus4) |  |  |  |  |  |
| Nairobi sheep disease virus4) |  |  |  |  |  |
| Ross river virus4) |  |  |  |  |  |
| Caprine arthritis encephalitis virus4) |  |  |  |  |  |
| Sheeppox virus4) |  |  |  |  |  |
| Peste des petits ruminants virus4) |  |  |  |  |  |
| Rinderpest virus4) |  |  |  |  |  |
| *Chlamydia abortus*4) |  |  |  |  |  |
| *Leptospira*4*)* |  |  |  |  |  |

Note : All health conditions and abnormalities of sheep shall be completely recorded. For dead sheep, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, SN: Serum neutralization test, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, AGP: Agar gel precipitation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

5) Absence of scrapie need to be demonstrated by a proof of the disease absence for at least two years and histological examination of brain materials from slaughtered animals and/or testing for scrapie-related myofibril.

Table 3 Inspection and Treatment of SPF Goat Flocks

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Rotavirus | Tochigi | Every three months | Either five animals per group or 10% of the animals, whichever is greater | SN | All antibody-positive group and cohabitant group 3) culling |
| Bovine adenovirus | Fukuroi | 〃 | 〃 | SN | 〃 |
| Akabane virus | JaGAr39 | 〃 | 〃 | SN | 〃 |
| Aujeszky's disease virus |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| Bluetongue virus |  | 〃 | 〃 | AGP | All the antibody-positive group and cohabitant group culling |
| Infectious bovine rhinotracheitis virus | No.758 | 〃 | 〃 | SN | 〃 |
| Parainfluenza virus type 3 | BN-1 | 〃 | 〃 | HI | 〃 |
| Caprine arthritis-encephalitis virus |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| Contagious pustular dermatitis virus |  | 〃 | 〃 | SN | All the antibody-positive group and cohabitant group culling |
| Visna-maedi virus |  | 〃 | 〃 | SN | 〃 |
| Japanese encephalitis virus | Nakayama | 〃 | 〃 | HI | 〃 |
| Malignant catarrhal fever virus | Cattle/elk type | 〃 | 〃 | IFA | 〃 |
| *Pasteurella multocida* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Mycoplasma* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Leptospira* |  | 〃 | 〃 | Clinical symptoms | 〃 |
| *Mycobacterium bovis, Mycobacterium tuberculosis* |  | 〃 | 〃 | Tuberculin reaction | All the antibody-positive group and cohabitant group culling |
| *Mycobacterium avium subsp. paratuberculosis* |  | 〃 | 〃 | Johnin reaction | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Brucella* |  | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| Epidemic hemorrhagic disease virus4) |  |  |  |  |  |
| Cowpox virus, Vaccinia virus4) |  |  |  |  |  |
| Foot and mouth disease virus4) |  |  |  |  |  |
| Rift valley fever virus4) |  |  |  |  |  |
| Caprine herpesvirus4) |  |  |  |  |  |
| Border disease virus4) |  |  |  |  |  |
| Peste des petits ruminants virus4) |  |  |  |  |  |
| Sheeppox virus4) |  |  |  |  |  |
| Rinderpest virus4) |  |  |  |  |  |
| Nairobi Sheep disease virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of goats shall be completely recorded. For dead goats, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, SN: Serum neutralization test, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, AGP: Agar gel precipitation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 4 Inspection and Treatment of SPF Horse Herds

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Equine rhinovirus |  | Every three months | Either five animals per group or 10% of the animals, whichever is greater | SN | All the antibody-positive group and cohabitant group 3) culling |
| Equine Getah virus | AMM-2021 Haruna | 〃 | 〃 | SN HI | 〃 |
| Equine herpesvirus | Type 1: Equine rhinopneumonitis virus HH-1BKS | 〃 | 〃 | ELISA | 〃 |
| Equine infectious anemia virus | P-337-EFD\*  P-337 | 〃 | 〃 | AGP ELISA | 〃 |
| Equine influenza virus |  | 〃 | 〃 | HI | 〃 |
| Japanese encephalitis virus | Nakayama | 〃 | 〃 | HI | 〃 |
| *Salmonella abortus equi* | *Salmonella abortus equi* | 〃 | 〃 | AGG | 〃 |
| *Clostridium tetani* |  | 〃 | 〃 | Clinical symptoms | All the positive group and cohabitant group culling |
| *Taylorella equigenitalis* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| Equine adenovirus4) |  |  |  |  |  |
| African horse sickness virus4) |  |  |  |  |  |
| Borna disease virus4) |  |  |  |  |  |
| West Nile virus4) |  |  |  |  |  |
| Equine viral arteritis virus4) |  |  |  |  |  |
| Equine encephalomyelitis virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |
| Vesicular stomatitis virus4) |  |  |  |  |  |
| Nipah virus4) |  |  |  |  |  |
| Hendra virus4) |  |  |  |  |  |
| Horsepox virus4) |  |  |  |  |  |
| *Burkholderia pseudomallei*4) |  |  |  |  |  |
| *Streptococcus equi subsp. equi*4) |  |  |  |  |  |
| *Trypanosoma*4) |  |  |  |  |  |

Note : All health conditions and abnormalities of horses shall be completely recorded. For dead horses, histopathological examination and other associated examinations should be performed.

1) The test antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, SN: Serum neutralization test, AGG: Agglutination test, AGP: Agar gel precipitation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 5 Inspection and Treatment of SPF Swine Herds

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Porcine adenovirus | Mie | Every three months | Either five animals per group or 10% of the animals, whichever is greater | SN | All antibody-positive group and cohabitant group 3) culling |
| Japanese encephalitis virus | Nakayama | 〃 | 〃 | HI | 〃 |
| Porcine Getah virus | Kanagawa, Haruna | 〃 | 〃 | HI ELISA | 〃 |
| Porcine circovirus |  | 〃 | 〃 | IFA | 〃 |
| Aujeszky's disease virus | Shope Sullivan | 〃 | 〃 | ELISA, latex agglutination | 〃 |
| Bovine viral diarrhea virus | Nose | 〃 | 〃 | SN | 〃 |
| Swine fever virus | ALD-A76 | 〃 | 〃 | ELISA | 〃 |
| Encephalomyocarditis virus |  | 〃 | 〃 | SN | 〃 |
| Porcine hemagglutinating encephalomyelitis virus | HEV-67 | 〃 | 〃 | SN | 〃 |
| Swine transmissible gastroenteritis virus | TO-K | 〃 | 〃 | SN | 〃 |
| Porcine cytomegalovirus |  | 〃 | 〃 | IFA | 〃 |
| Porcine epidemic diarrhea virus |  | 〃 | 〃 | SN | 〃 |
| Porcine enterovirus (excluding porcine *Teschovirus*) |  | 〃 | 〃 | SN (A: PEV-9UKG/410/73, B: PEV-8V13) | 〃 |
| Porcine influenza virus |  | 〃 | 〃 | HI (H1N1, H1N2, H3N2) | 〃 |
| Porcine parvovirus | 90HS-SK | 〃 | 〃 | HI | 〃 |
| Porcine reproductive and respiratory syndrome virus | Lelystad | 〃 | 〃 | ELISA | 〃 |
| Porcine rotavirus | S-80 | 〃 | 〃 | SN | 〃 |
| *Mycoplasma* | *hyopneumoniae* | 〃 | 〃 | ELISA | 〃 |
| *Brachyspira hyodysenteriae* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Lawsonia intracellularis* |  | 〃 | 〃 | Clinical symptoms | 〃 |
| *Mycobacterium avium-intracellulare* | *Mycobacterium avium* | 〃 | 〃 | Tuberculin reaction | All the antibody-positive group and cohabitant group culling |
| *Actinobacillus pleuropneumoniae* | Serotype2 SHP-1 | 〃 | 〃 | AGG | 〃 |
| *Bordetella bronchiseptica* | H-16 | 〃 | 〃 | AGG | 〃 |
| *Brucella* | *Melitensis* | 〃 | 〃 | CF | 〃 |
| *Erysipelothrix rhusiopathiae* | Tama 96 | 〃 | 〃 | Latex agglutination | 〃 |
| *Pasteurella multocida* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Leptospira* |  | 〃 | 〃 | Clinical symptoms | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Toxoplasma* |  | 〃 | 〃 | Color tone test | 〃 |
| Porcine teschovirus4) |  |  |  |  |  |
| African swine fever virus4) |  |  |  |  |  |
| Foot and mouth disease virus4) |  |  |  |  |  |
| Porcine herpesvirus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |
| Swinepox virus4) |  |  |  |  |  |
| Swine vesicular disease virus4) |  |  |  |  |  |
| Vesicular stomatitis virus4) |  |  |  |  |  |
| Rinderpest virus4) |  |  |  |  |  |
| Nipah virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of swine shall be completely recorded. For dead swine, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, SN: Serum neutralization test, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, AGP: Agar gel precipitation test , CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 6 Inspection and Treatment of SPF Dog Groups

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Aujeszky's disease virus |  | Every three months | Either five breeding animals per group or 10% of the animals, whichever is greater | Clinical symptoms | All the positive group and cohabitant group3)culling |
| Canine adenovirus |  | 〃 | 〃 | SN | All the antibody-positive group and cohabitant group culling |
| Canine coronavirus |  | 〃 | 〃 | SN | 〃 |
| Canine distemper virus |  | 〃 | 〃 | SN | 〃 |
| Canine herpesvirus |  | 〃 | 〃 | SN | 〃 |
| Canine parvovirus |  | 〃 | 〃 | HI | 〃 |
| Canine parainfluenza virus |  | 〃 | 〃 | SN | 〃 |
| *Brucella canis* | Canis | 〃 | 〃 | AGG | 〃 |
| *Leptospira* |  | 〃 | 〃 | AGG | 〃 |
| Rabies virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of dogs shall be completely recorded. For dead dogs, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, SN: Serum neutralization test, AGG: Agglutination test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 7 Inspection and Treatment of SPF Cat Groups

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Aujeszky's disease virus |  | Every three months | Either five breeding animals per group or 10% of the animals, whichever is greater | Clinical symptoms | All the positive group and cohabitant group3) culling |
| Feline calicivirus |  | 〃 | 〃 | SN | All the antibody-positive group and cohabitant group culling |
| Feline herpesvirus type 1 |  | 〃 | 〃 | SN | 〃 |
| Feline immunodeficiency virus |  | 〃 | 〃 | ELISA | 〃 |
| Feline leukemia virus/Feline sarcoma virus |  | 〃 | 〃 | Colloidal gold-labeled antibody chromatography | 〃 |
| Feline infectious peritonitis virus/Feline enteric coronavirus |  | 〃 | 〃 | ELISA | 〃 |
| Feline panleukopenia virus |  | 〃 | 〃 | HI | 〃 |
| Feline syncytial virus |  | 〃 | 〃 | SN | 〃 |
| *Chlamydophila felis* |  | 〃 | 〃 | CF | 〃 |
| Cowpox virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of cats shall be completely recorded. For dead cats, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, SN: Serum neutralization test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 8 Inspection and Treatment of SPF Rabbit Colonies

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Sendai virus | MN | Every one month | Random sampling of ten from retired animals | CF | All the antibody-positive group and cohabitant group 3) culling |
| Japanese encephalitis virus |  | 〃 | 〃 | HI | 〃 |
| *Mycoplasma pulmonis* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Pseudomonas aeruginosa* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Typhimurium | K-28 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Pasteurella pneumotropica* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Pasteurella multocida* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Bordetella bronchiseptica* | 64L | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Streptococcus pneumoniae* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Streptococcus zooepidemicus* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Bacillus piliformis* | RT MSK | 〃 | 〃 | CF ELISA IFA | All the antibody-positive group and cohabitant group culling |
| *Eimeria* |  | 〃 | 〃 | Microscopic examination | All the positive group and cohabitant group culling |
| *Psoroptes cuniculi* |  | 〃 | 〃 | Microscopic examination | 〃 |
| Rabbitpox virus |  | 〃 | 〃 | ELISA IFA | All the antibody-positive group and cohabitant group culling |
| Rabies virus4) |  |  |  |  |  |
| Myxoma virus/Rabbit fibroma virus4) |  |  |  |  |  |
| Rabbit hemorrhagic disease virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of rabbits shall be completely recorded. For dead rabbits, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, IFA: Indirect immunofluorescence test, AGG: Agglutination test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 9 Inspection and Treatment of SPF Mouse Colonies

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Sendai virus | MN | Every one month | Random sampling of ten from retired animals | CF | All the antibody-positive group and cohabitant group 3) culling |
| Pneumoniae virus of mice |  | 〃 | 〃 | ELISA IFA | 〃 |
| Mouse hepatitis virus | Pr | 〃 | 〃 | CF | 〃 |
| Lymphocytic choriomeningitis virus |  | 〃 | 〃 | ELISA IFA | 〃 |
| Ectromelia virus |  | 〃 | 〃 | ELISA IFA | 〃 |
| *Mycoplasma pulmonis* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Pseudomonas aeruginosa* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Citrobacter rodentium* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Typhimurium | K-28 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Pasteurella pneumotropica* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Corynebacterium kutscheri* | CK-1 | 〃 | 〃 | Bacterial isolation | 〃 |
|  |  | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Bacillus piliformis* | MSK RT | 〃 | 〃 | ELISA IFA | 〃 |
| *Helicobacter hepaticus* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Giardia muris* |  | 〃 | 〃 | Microscopic examination | 〃 |
| *Spironucleus miris* |  | 〃 | 〃 | Microscopic examination | 〃 |
| *Syphacia obvelata* |  | 〃 | 〃 | Microscopic examination | 〃 |
| *Escherichia coli* | *E.coli Ol15 a,c:K (B)* | 〃 | 〃 | Bacterial isolation | 〃 |
| Dermatophytes |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Enteritidis | EC-5 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Hexamita muris* |  | 〃 | 〃 | Microscopic examination | All the positive group and cohabitant group culling |
| *Trichomonas muris* |  | 〃 | 〃 | Microscopic examination | Update the original group if positive |
| *Entamoeba muris* |  | 〃 | 〃 | Microscopic examination | 〃 |
| Encephalomyocarditis virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |
| Mouse adenovirus4) |  |  |  |  |  |
| Mouse rotavirus4) |  |  |  |  |  |
| Reovirus4) |  |  |  |  |  |
| Mouse parvovirus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of mice shall be completely recorded. For dead mice, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. ELISA: Enzyme-linked immunosorbent assay, IFA: Indirect immunofluorescence test, AGG: Agglutination test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 10 Inspection and Treatment of SPF Rat Colonies

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Sialodacryoadenitis virus |  | Every one month | Random sampling of ten from retired animals | CF | All the antibody-positive group and cohabitant group 3) culling |
| Sendai virus | MN | 〃 | 〃 | CF | 〃 |
| Hantavirus |  | 〃 | 〃 | ELISA IFA | 〃 |
| Pneumoniae virus of mice |  | 〃 | 〃 | ELISA IFA | 〃 |
| *Mycoplasma pulmonis* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Pseudomonas aeruginosa* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Typhimurium | K-28 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Pasteurella pneumotropica* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Bordetella bronchiseptica* |  | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Streptococcus pneumoniae* | 64L | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Corynebacterium kutscheri* | CK-1 | 〃 | 〃 | Bacterial isolation | 〃 |
| *Bacillus piliformis* |  | 〃 | 〃 | ELISA IFA | All the antibody-positive group and cohabitant group culling |
| *Giardia muris* |  | 〃 | 〃 | Microscopic examination | All the positive group and cohabitant group culling |
| *Spironucleus miris* |  | 〃 | 〃 | Microscopic examination | 〃 |
| *Syphacia obvelata* |  | 〃 | 〃 | Microscopic examination | 〃 |
| Mouse hepatitis virus | Pr | 〃 | 〃 | CF | All the antibody-positive group and cohabitant group culling |
| Dermatophytes |  | 〃 | 〃 | Bacterial isolation | 〃 |
| Mouse adenovirus4) |  |  |  |  |  |
| Encephalomyocarditis virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |
| Mouse rotavirus4) |  |  |  |  |  |
| Reovirus4) |  |  |  |  |  |
| Rat parvovirus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of rats shall be completely recorded. For dead rats, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. ELISA: Enzyme-linked immunosorbent assay, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

# Table 11 Inspection and Treatment of SPF Hamster Colonies

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Sendai virus | MN | Every one month | Random sampling of ten from retired animals | CF | All the antibody-positive group and cohabitant group 3) culling |
| Hantavirus |  | 〃 | 〃 | ELISA IFA | 〃 |
| Pneumoniae virus of mice |  | 〃 | 〃 | ELISA IFA | 〃 |
| Lymphocytic choriomeningitis virus |  | 〃 | 〃 | ELISA IFA | 〃 |
| *Mycoplasma pulmonis* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Pseudomonas aeruginosa* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Typhimurium | K-28 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Pasteurella pneumotropica* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Bordetella bronchiseptica* |  | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Streptococcus pneumoniae* | 64L | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Corynebacterium kutscheri* | CK-1 | 〃 | 〃 | Bacterial isolation | 〃 |
| *Bacillus piliformis* |  | 〃 | 〃 | ELISA IFA | All the antibody-positive group and cohabitant group culling |
| *Giardia muris* |  | 〃 | 〃 | Microscopic examination | All the positive group and cohabitant group culling |
| *Spironucleus miris* |  | 〃 | 〃 | Microscopic examination | 〃 |
| *Syphacia obvelata* |  | 〃 | 〃 | Microscopic examination | 〃 |
| *Yercinia pseudotuberclosis* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| Encephalomyocarditis virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of hamsters shall be completely recorded. For dead hamsters, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. ELISA: Enzyme-linked immunosorbent assay test, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.

Table 12 Inspection and Treatment of SPF Guinea pig Colonies

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Pathogen | Antigen under inspection1) | Inspection timing and number of animals inspected | | Inspection method2) | Treatment |
| Timing | Number of animals |
| Sendai virus | MN | Every one month | Random sampling of ten from retired animals | CF | All the antibody-positive group and cohabitant group 3) culling |
| Japanese encephalitis virus |  | 〃 | 〃 | HI | 〃 |
| *Mycoplasma caviae* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Mycoplasma pulmonis* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Pseudomonas aeruginosa* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Typhimurium | K-28 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Pasteurella pneumotropica* |  | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Bordetella bronchiseptica* |  | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Streptococcus pneumoniae* | 64L | 〃 | 〃 | Bacterial isolation | All the positive group and cohabitant group culling |
| *Streptococcus zooepidemicus* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Bacillus piliformis* |  | 〃 | 〃 | ELISA IFA | All the antibody-positive group and cohabitant group culling |
| *Eimeria* |  | 〃 | 〃 | Microscopic examination | All the positive group and cohabitant group culling |
| *Psoroptes cuniculi* |  | 〃 | 〃 | Microscopic examination | 〃 |
| Dermatophytes |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Yercinia pseudotuberculosis* |  | 〃 | 〃 | Bacterial isolation | 〃 |
| *Salmonella* Enteritidis | EC-5 | 〃 | 〃 | AGG | All the antibody-positive group and cohabitant group culling |
| *Trichomonas muris* |  | 〃 | 〃 | Microscopic examination | Update the original group if positive |
| Encephalomyocarditis virus4) |  |  |  |  |  |
| Rabies virus4) |  |  |  |  |  |

Note : All health conditions and abnormalities of guinea pigs shall be completely recorded. For dead guinea pigs, histopathological examination and other associated examinations should be performed.

1) The antigen may be substituted with another appropriate strain.

2) An equivalent inspection method may be applied, if available. The alternate inspection method to be applied shall be verified and guaranteed to ensure their validity. HI: Hemagglutination inhibition test, ELISA: Enzyme-linked immunosorbent assay, IFA: Indirect fluorescence antibody test, AGG: Agglutination test, CF: Complement fixation test

3) Cohabitant group refers to a group of animals not completely isolated from the positive group.

4) For pathogens without reported field case in Japan (or those deemed less severe), the selection of antigens, inspection methods, and treatments should prioritize those applied in the countries reported.